

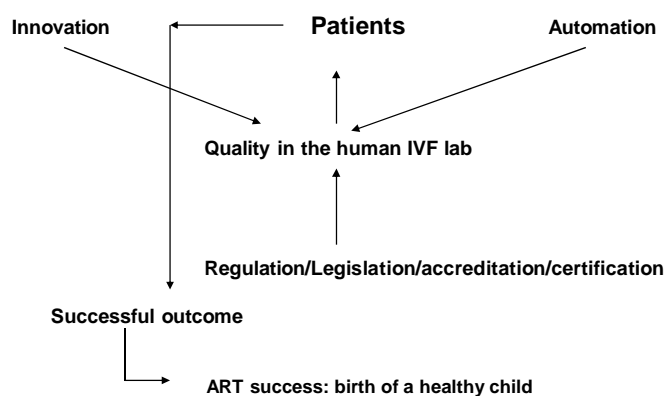
## Blastocyst culture and transfer: a tool to improve the efficiency of ART and to reduce multiple pregnancies?

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Department of Reproductive Medicine, University Hospital Gent, Belgium  
KLEM 2014


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### Introduction



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The IVF lab and the challenges of the 21st century

Improving the efficiency of ART(per oocyte collection cycle) - avoiding multiple pregnancies

Single ET and Single FRET

Optimal embryo selection for transfer and cryopreservation


- Which, how, when and why?


Two questions:

- How many healthy live births are obtained from one stimulation cycle?
- How many stimulation cycles are necessary for a patient to give birth to one healthy baby?

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ART success: birth of one healthy child


Transfer of one embryo: SET and sFRET

Gardner D et al (1998) Culture and transfer of human blastocysts increases implantation rates and reduces the need for multiple embryo transfers. Fertil Steril 69, 84-88.


The road for single embryo transfer = blastocyst transfer?

Yes or No

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**ART success: birth of one healthy child**  
**Transfer of one embryo: SET and sFRET**


Day of embryo transfer  
 Benefit of blastocyst transfer  
 =  
 Subject of debate

**2014?**

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**Rationale for blastocyst culture and ET**

- ➔ Synchronization of embryo and uterine development
- ➔ Provide a mechanism of self-selection of the viable embryos

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### Rationale for blastocyst culture and ET

- Synchronization of embryo and uterine development

Bovine: blastocyst transfer is routine, highly successful and the only viable option because cleaved embryos cannot survive the bovine uterine environment (Thompson and Peterson, 2000)

Primates: pronucleate and early cleaving embryos can survive in the uterine environment (Martson et al, 1977)

Human: First IVF pregnancy = blastocyst transfer, blastocyst transfer was not universally adopted


Transfer on day 0, day 1, day 2, day 3, day 4, ...occasionally day 5/6


Since 1998: transfer on day 2/3 and also on day 5, day 6 ...

Current debate in the human

Comparison between uterine transfer of cleavage-stage and blastocyst stage embryos

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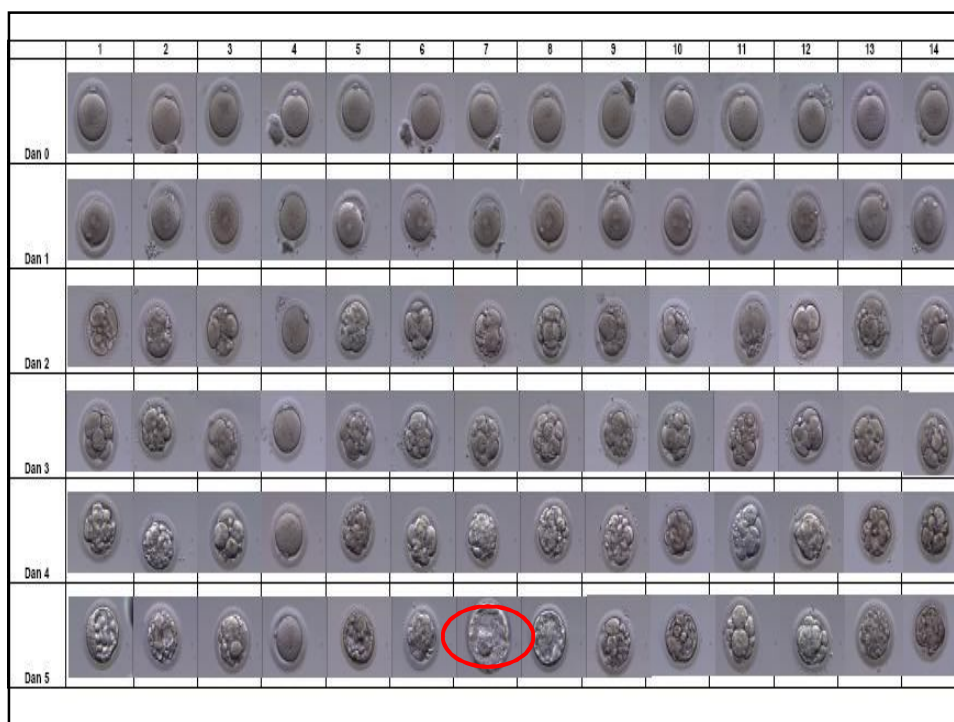

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### Rationale for blastocyst culture and ET

- Provide a mechanism of self-selection of the viable embryos


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**Rationale for blastocyst culture and ET**

- ➔ Provide a mechanism of self-selection of the viable embryos

Adler et al (2013) Blastocyst culture selects for euploid embryos: comparison of blastomere and trophectoderm biopsies. RBM Online , Article in press

- ➔ aCGH analysis to assess all 24 chromosomes
- ➔ Day 3 blastomeres and day 5 trophectoderm
- ➔ 1603 embryos
- ➔ A significant larger proportion of embryos were euploid on day 5 biopsy (42%) compared with day 3 biopsy (24%)

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**Can we select embryos with a high developmental potential earlier in development for transfer rather than wait until they develop to the blastocyst stage?**

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Megaset study: objectives and methods (Van den Abbeel et al, RBM Online, 2013, Ziebe et al, Fertil Steril, 2012))

**Objective**

- To correlate embryo quality on day 3 and 4 with blastocyst development on day 5
- Investigate the relationship between blastocyst quality and pregnancy in a SET trial

**Design**


- MEGASET: Randomized, assessor-blind, parallel group, multicentre trial comparing HP-hMG with rFSH in GnRH antagonist cycles (N=749)


**Methods**

- Central training meetings and scoring sessions for all embryologists, and reference atlas
- Quality assessments at fixed, narrow time-points
- All oocytes were inseminated using ICSI and cultured to blastocyst stage ( $120 \pm 2$  hours)
- Compulsory SET of a blastocyst on Day 5 for all patients (N=618)
- All supernumerary blastocysts were vitrified
- Pregnancy monitoring:
  - Serum  $\beta$ -hCG (13–15 days after ET)
  - Clinical pregnancy (5–6 weeks after ET)
  - Ongoing pregnancy (10–11 weeks after ET)

Early pregnancy loss  
= positive  $\beta$ -hCG, but no ongoing pregnancy

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## Relationship: Day 3 → Day 5

| Day 3           | Day 5                            |                             |                              |                    |
|-----------------|----------------------------------|-----------------------------|------------------------------|--------------------|
|                 | Excellent-quality blastocyst (%) | Good-quality blastocyst (%) | Other-quality blastocyst (%) | Not blastocyst (%) |
| Top-quality     | 22                               | 24                          | 27                           | 27                 |
| Good-quality    | 10                               | 14                          | 28                           | 47                 |
| Minimum-quality | <1                               | 6                           | 17                           | 76                 |

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## Definitions of Embryo Quality on Day 3

| Top-quality embryo*   | Good-quality embryo*  | Minimum-quality embryo*   |
|---|---|---|
| <ul style="list-style-type: none"> <li>⚡ 4 or 5 blastomeres (day 2)</li> <li>⚡ 7 blastomeres (day 3)</li> <li>⚡ 20% fragmentation (day 3)</li> <li>⚡ equally sized blastomeres (day 3)</li> <li>⚡ no sign of multinucleation (days 1, 2 and 3)</li> </ul> | <ul style="list-style-type: none"> <li>⚡ 6 blastomeres (day 3)</li> <li>⚡ no cleavage arrest (i.e. cleavage must have occurred within the last 24h)</li> <li>⚡ 30% fragmentation (day 3)</li> <li>⚡ no sign of multinucleation (days 1, 2 and 3)</li> </ul> | <ul style="list-style-type: none"> <li>⚡ 4 blastomeres (day 3)</li> <li>⚡ no cleavage arrest (i.e. cleavage must have occurred within the last 24h)</li> <li>⚡ 50% fragmentation (day 3)</li> <li>⚡ no sign of multinucleation (days 1, 2 and 3)</li> </ul> |

\* all criteria must be fulfilled

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## **More observations is better embryo selection: Time Lapse imaging**

### Limitations of Time lapse systems

- Morphokinetic markers different from lab to lab
- TLS does not allow rotation of embryos making visual observations difficult
- Morphokinetic markers algorithms work with quartiles or time range which create very exact and sharp limits and this may not simulate embryo development.
- Morphokinetic markers up to the five cell stage is questionable since embryo development up to that stage is governed by maternal influences only
- Further requirements for innovations in both optics and image analysis
- Can a single culture medium without renewal on day 3 be developed which supports development throughout the pre-implantation period?

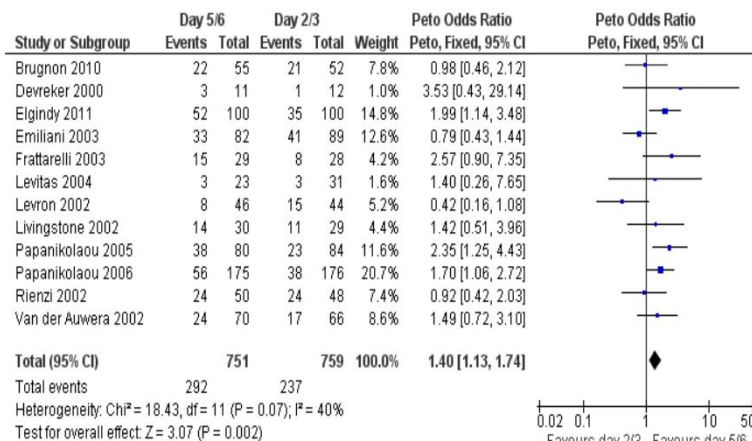
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**What are we trying to achieve with blastocyst transfer? Increased live birth rates, decreased multiple gestations or both?**

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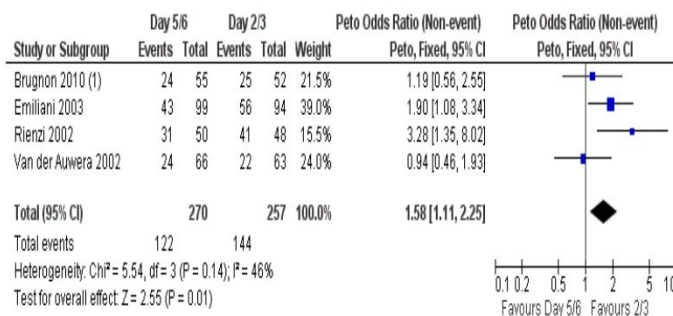


### Live birth rate per couple: Cleavage&Blastocyst transfer (Glujovski et al, 2012, Cochrane review, following on Blake 2004, 2005, 2007)




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### Cumulative fresh and frozen



(1) Study had policy of single embryo transfer

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### Authors' conclusions

- This review provides evidence that there is a *small* significant difference in live birth rates in favour of blastocyst transfer (Day 5 to 6) compared to cleavage stage transfer (Day 2 to 3). However, cumulative clinical pregnancy rates from cleavage stage (derived from fresh and thaw cycles) resulted in higher clinical pregnancy rates than from blastocyst cycles. The most likely explanation for this is the higher rates of frozen embryos and lower failure to transfer rates per couple obtained from cleavage stage protocols.

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## Cochrane review *Discussion*


**Considerations on Culture media used**

**Large differences between trials**

- Sequential media for blastocyst culture (14 trials)
- Sequential media for both arms (7 trials)
- Different commercial media for the two arms
- In-house made formulations for early embryos

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## Cochrane review *Discussion*

**Considerations on** Differences in lab policies

- Culture media used
- Culture conditions (must be optimal)
- Minimum criteria for embryo quality for ET/freezing
  - ⇒ ET cancellation rate / embryo freezing rate
- Methodology
- Blastulation rates
- Number of embryos transferred

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Papanikolaou et al. (*New Engl J Med* 354, 1139, 2006)

- July 2003 – Nov 2004
- <36 years
- Rank 1 or 2, IVF or ICSI
- Cycles planned for **SET**
- Inclusion and randomisation at consultation

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Papanikolaou et al. (*New Engl J Med* 354, 1139, 2006)

|                         | D3 ET<br>n=171 | D5 ET<br>n=169 | statistics |
|-------------------------|----------------|----------------|------------|
| ET rate (%)             | 157 (91.8)     | 150 (88.8)     | NS         |
| Clinical PR (%)         | 41 (24.0)      | 58 (34.3)      | P=0.04     |
| Ongoing PR (%)          | 38 (22.2)      | 58 (34.3)      | P=0.01     |
| Delivery rate (%)       | 38 (22.2)      | 56 (33.1)      | P=0.03     |
| Mult. delivery rate (%) | 2 (5.2)*       | 0 (0.0)        | NS         |

\*2 monozygotic twins

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## Further questions

- **Blastocyst grading:** Gardner's criteria are based on many subjective observations. Do we need to modify/modulate the system further?
- **Cryopreservation of blastocysts:** will vitrication methods change the picture?
- **Culture media:** how many different formulations do we need? Are further refinements necessary?

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## Blastocyst grading: Gardner's criteria are based on many subjective observations. Do we need to modify/modulate the system further?

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## Blastocyst classification

**Blastocyst expansion and hatching status**

1. An early blastocyst with a blastocoel being less than half the volume of that of the embryo
2. A blastocyst with a blastocoel volume half that, or greater than half that, of the embryo
3. A blastocyst with a blastocoel completely filling the embryo
4. An expanded blastocyst with a blastocoel volume larger than that of the early embryo, with a thinning zona
5. A hatching blastocyst with the trophectoderm starting to herniate through the zona
6. A hatched blastocyst, in which the blastocyst has completely escaped from the zona

For blastocysts with expansion and hatching status 3-6, blastocyst inner cell mass grading and trophectoderm grading were evaluated.

**Inner cell mass**


- A. Tightly packed, many cells
- B. Loosely grouped, several cells
- C. Very few cells


**Trophectoderm**

- A. Many cells forming a cohesive epithelium
- B. Few cells forming a loose epithelium
- C. Very few, large cells

Gardner DK, et al (eds). Towards reproductive certainty: infertility and genetics beyond. Camforth: Parthenon Press;1999:376-388

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## Literature

- Blastocyst quality considering composite scores has been linked to pregnancy and/or pregnancy loss (Gardner & Schoolcraft, 1999; Balaban et al, 2000, 2006; Gardner et al, 2000, 2004; Kovacic et al, 2008; Alpha Scientists / ESHRE SIG Embryology, 2011)
- Blastocyst quality: individual parameters vs composite score (Dokras et al, 1993; Balaban et al, 2000; Shapiro et al, 2000, 2008; Richter et al, 2001; Ahlström et al, 2011, Hill et al, 2013)
- Most of the data in the literature are from multiple embryo transfers, and there are limited data on SET (Ahlström et al, 2011, Hill et al, 2013)

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## Megaset study: objectives and methods

### Objective

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### Design

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Early pregnancy loss  
= positive  $\beta$ -hCG, but no ongoing pregnancy

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### Logistic regression models

- Identification of statistically significant ( $p < 0.05$ ) predictors of pregnancy and early pregnancy loss**

Approach: Testing of one parameter at a time

Approach: Model including all parameters (expansion and hatching status, inner cell mass, trophectoderm)

| Outcome               | INDIVIDUAL ANALYSIS Significant predictors  | COMBINED MODEL Significant predictors   |
|-----------------------|---|---|
| Positive $\beta$ -hCG | <ul style="list-style-type: none"> <li>Expansion and hatching status</li> <li>Inner cell mass</li> <li>Trophectoderm</li> </ul> | <ul style="list-style-type: none"> <li>Expansion and hatching status</li> </ul> |
| Ongoing pregnancy     | <ul style="list-style-type: none"> <li>Expansion and hatching status</li> <li>Inner cell mass</li> <li>Trophectoderm</li> </ul> | <ul style="list-style-type: none"> <li>Expansion and hatching status</li> </ul> |
| Early pregnancy loss  | <ul style="list-style-type: none"> <li>Inner cell mass</li> </ul>   | None  |

Van den Abbeel E et al. RBM Online, 2013

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### Predicted ongoing pregnancy rate

| Blastocyst and expansion | Inner cell mass | Trophectoderm | Ongoing pregnancy (%) |
|--------------------------|-----------------|---------------|-----------------------|
| 1                        | -               | -             | 6                     |
| 2                        | -               | -             | 12                    |
| 3                        | A               | A             | 25                    |
| 3                        | A               | B             | 22                    |
| 3                        | A               | C             | 21                    |
| 3                        | B               | A             | 20                    |
| 3                        | B               | B             | 18                    |
| 3                        | B               | C             | 17                    |
| 3                        | C               | A             | 16                    |
| 3                        | C               | B             | 14                    |
| 3                        | C               | C             | 14                    |
| 4-5                      | A               | A             | 48                    |
| 4-5                      | A               | B             | 45                    |
| 4-5                      | A               | C             | 44                    |
| 4-5                      | B               | A             | 42                    |
| 4-5                      | B               | B             | 38                    |
| 4-5                      | B               | C             | 37                    |
| 4-5                      | C               | A             | 35                    |
| 4-5                      | C               | B             | 32                    |
| 4-5                      | C               | C             | 31                    |


For blastocyst Stages 3 versus 4-5, selection should be based on blastocyst expansion and hatching status

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## Cryopreservation of blastocysts: will vitrification methods change the picture?

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### Which method to use: slow freezing or vitrification?

- There are two circumstances in which vitrification methods should definitively be considered: where it is clear that extra-cellular ice is responsible for significant damage, and where the results of classical freezing methods are unsatisfactory (Pegg, 2005)
- Classical freezing of blastocysts is suboptimal (Kolibianakis et al, 2010)

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### Results from literature: blastocysts (open vitrification)

Choi (2000), Yokota (2001), Cho (2002), Reed (2002), Hiraoka (2004), Hu (2004), Stehlik (2005), Huang (2005), Kuwayama (2005), Mukaida (2007), Liebermann (2007), Son (2007), Van der Zwalmen (2007), Hiraoka (2008), Ebner (2009) Liebermann (2009), Rama Raju (2009)

| Clin P / ET | Impl /E Transferred | Impl / E Warmed |
|-------------|---------------------|-----------------|
| 4974/10197  | 3124/11117          | 3124/13629      |
| (48.8%)     | (28.1%)             | (22.9%)         |

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### Results from literature: blastocysts (closed vitrification)

Stachecki (2008), Van der Zwalmen (2009, 2013), [Liebermann \(2009\)](#), [Van Landuyt \(2011\)](#), [De Croo \(2013\)](#) (= CBS HS VIT)

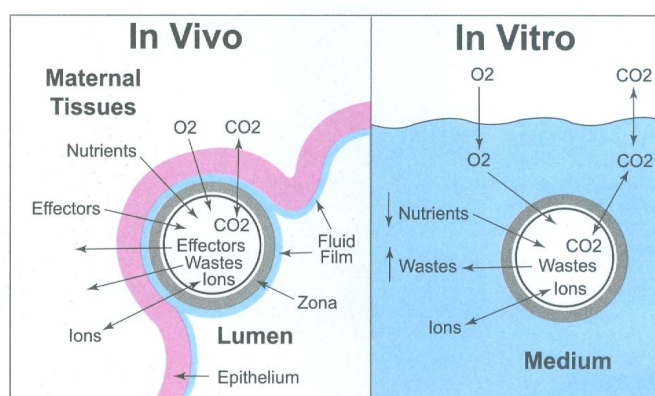
| Clin P / ET | Impl /E Transferred | Impl / E Warmed |
|-------------|---------------------|-----------------|
| 229/435     | 263/854             | 263/1004        |
| (52.6%)     | (30.8%)             | (26.2%)         |

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## Culture media: how many different formulations do we need? Are further refinements necessary?

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## Embryo culture



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Figure 1. A comparison of the interaction of an embryo with its environment, in vivo and in vitro.

## Composition of culture medium: debate

### Questions

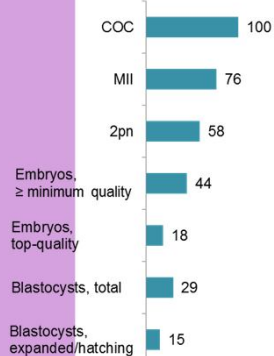
- ❖ Simple basic media used for day 2/3 culture are suboptimal for blastocyst culture?
- ❖ Should the composition of the medium be changed to correspond with the physiological changes which occur as development proceeds?
- ❖ Can a single culture medium be developed which supports development throughout the pre-implantation period?

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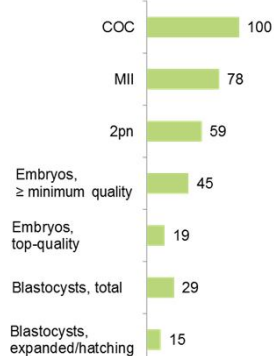
### Embryo development: MEGASET STUDY

#### Overall development from COC to blastocyst

##### HP-hMG Percentage



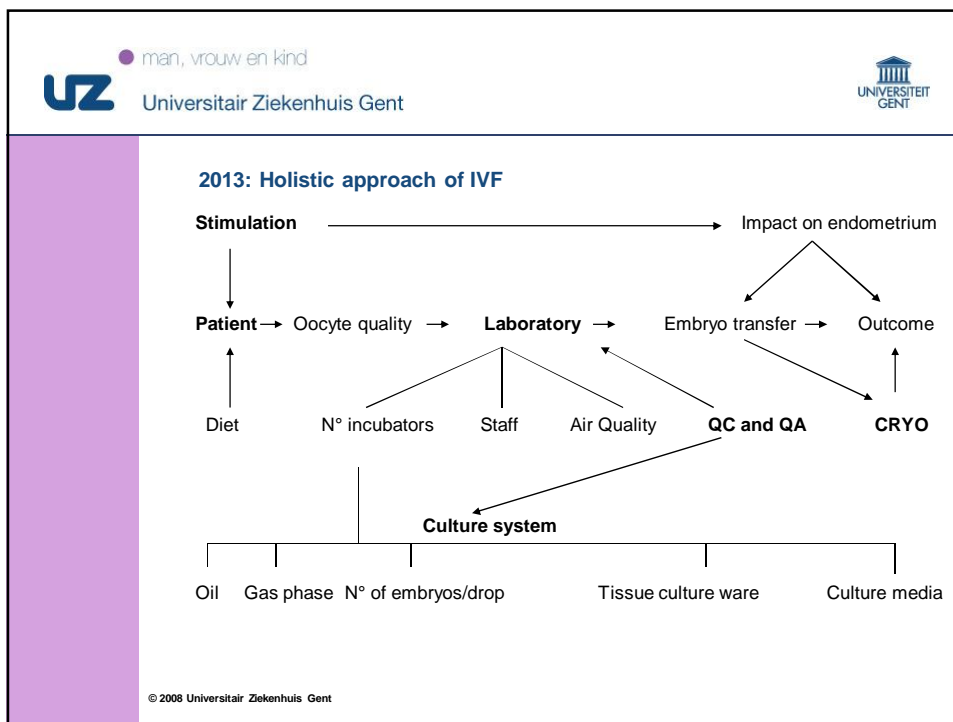
##### rFSH Percentage



#### Blastocyst expansion and hatching status

|   | HP-hMG | rFSH |
|---|--------|------|
| 1 | 8%     | 10%  |
| 2 | 9%     | 8%   |
| 3 | 16%    | 18%  |
| 4 | 41%    | 42%  |
| 5 | 26%    | 23%  |
| 6 | 0%     | 0%   |

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### Additional Considerations

- **Higher cost related to blastocyst culture – transfer**
  - Additional incubators
  - Optimized laboratory environment
  - Additional culture media
  - More laboratory staff members
  - Cost – benefit analysis ?
- **Can patients and ARTologists accept the psychological trauma of no ET if blastocysts do not develop in vitro?**

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The IVF lab and the challenges of the 21st century  
Single embryo transfer avoiding multiple pregnancies

- Ecological clinical embryology?
  - Transfer embryos early D2/D3
  - Embryo Selection?
- Blastocyst transfer?
  - Benefits of blastocyst culture and ET
  - Risks of long term culture?

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- **Blastocyst culture and single blastocyst transfer is a powerful tool to improve the efficiency of ART and to reduce multiple pregnancies**
- **Vitrification of blastocysts works and should be regarded as an established technique**
- **Time for an optimized RCT?**

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### The IVF lab and the challenges of the 21st century Single embryo transfer avoiding multiple pregnancies

- Ecological clinical embryology?
  - Transfer embryos early D2/D3
  - Embryo Selection?
- Blastocyst transfer?
  - Benefits of blastocyst culture and ET
  - Risks of long term culture?
- Freeze all but at the blastocyst stage?
  - Transfer in natural cycle
  - Transfer all embryos one at the time
  - No embryo selection
  - Optimized cryopreservation
  - Time to pregnancy?

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