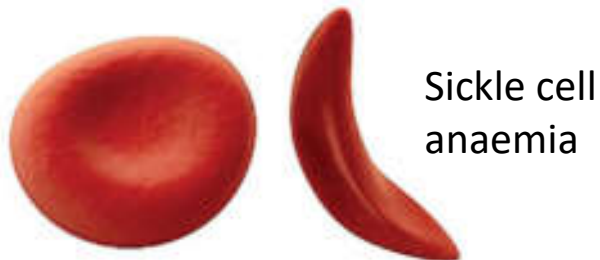


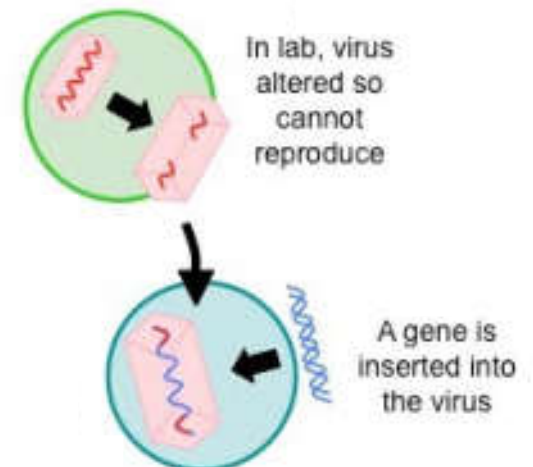
*An update on the worldwide controversy
on germline gene editing*

Dr. Ben Davies

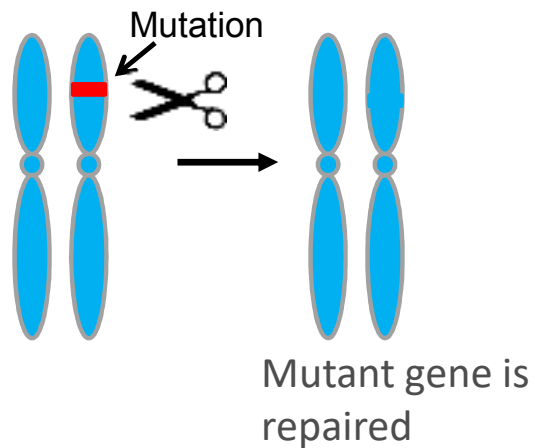
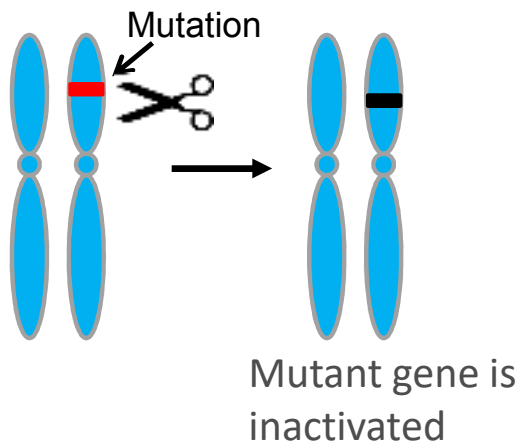
Gene Therapy - replacing a broken gene



- Mutation in a gene encoding one of the proteins that make up haemoglobin
- Currently gene therapy is focussed on delivering extra copies of healthy genes to patients with mutations

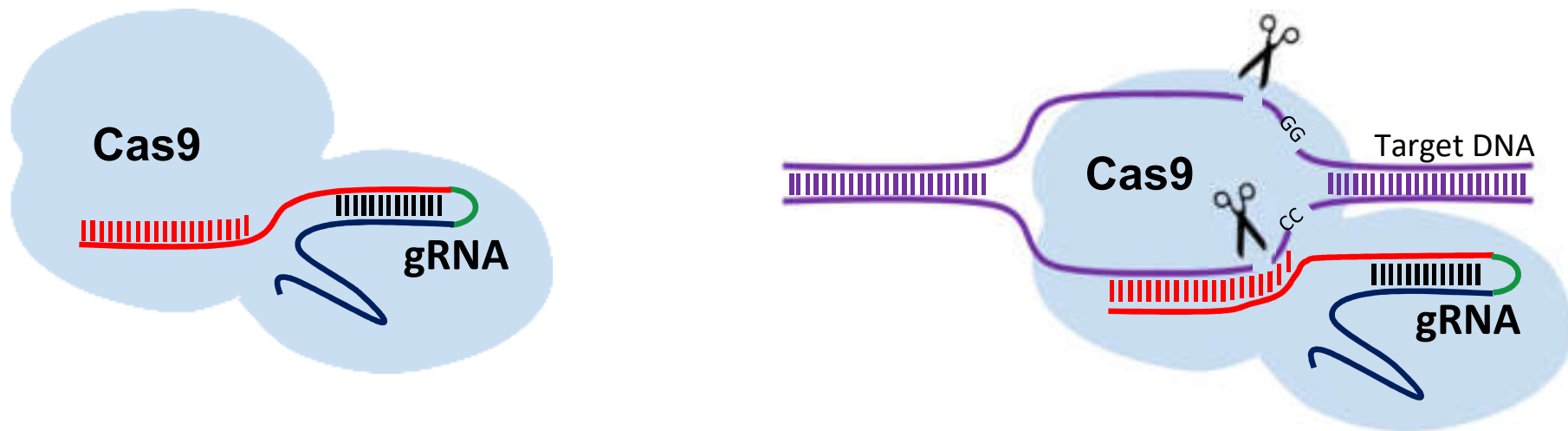


Could repairing the actual diseased genes be a better strategy?

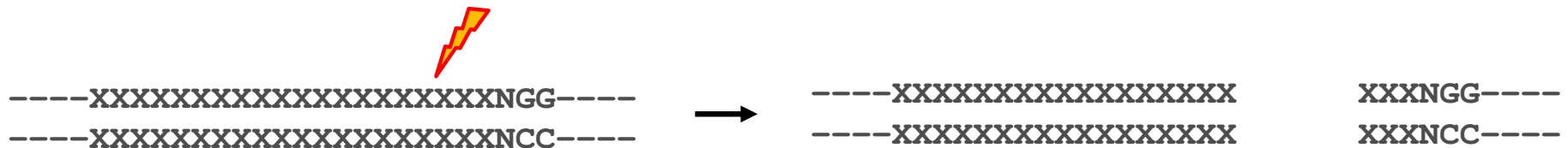


- So we need some molecular scissors
- This was, until recently, science fiction!

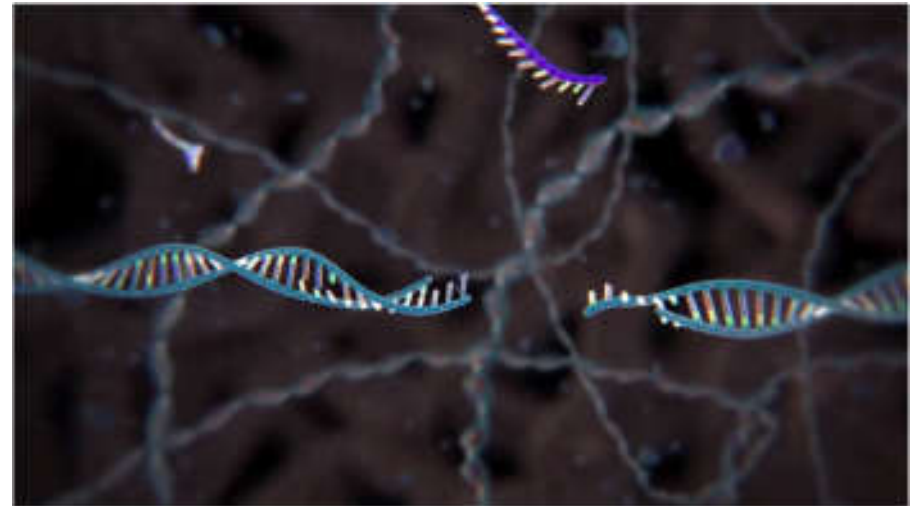
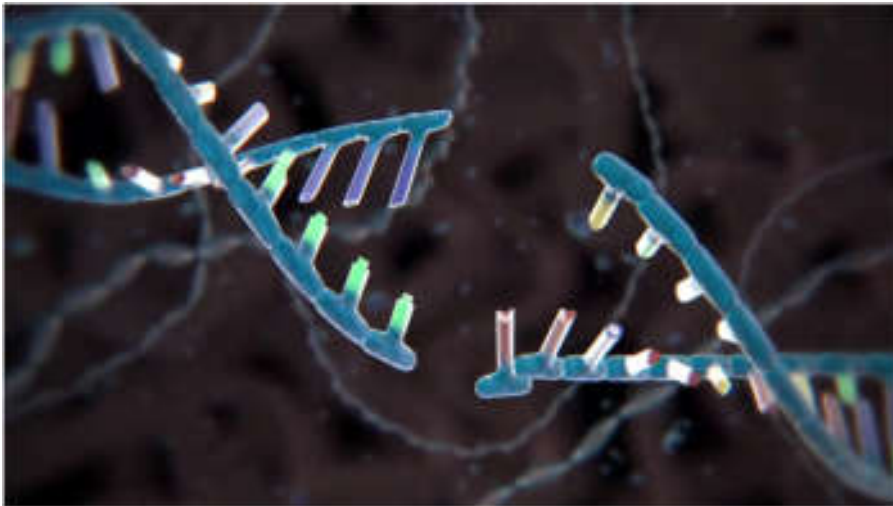
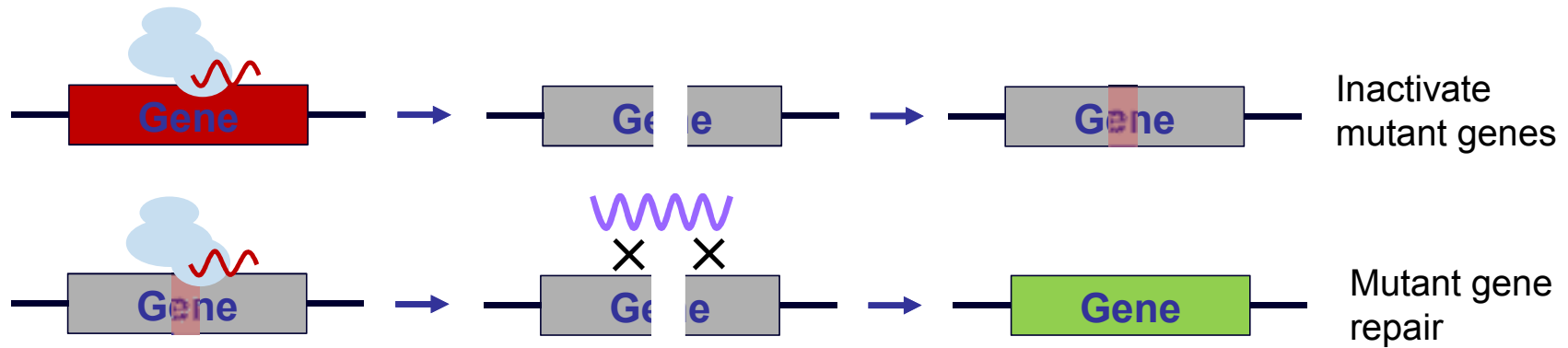
The Present – CRISPR/Cas9 (Clustered Regularly Interspaced Short Palindromic Repeats)



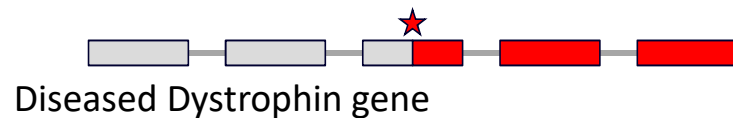
- Cas9 is the nuclease which cleaves its double stranded target site
- A guide-RNA (crRNA:tracrRNA) defines where the nuclease cleaves
- The target site can be any 20 nt sequence followed by -NGG



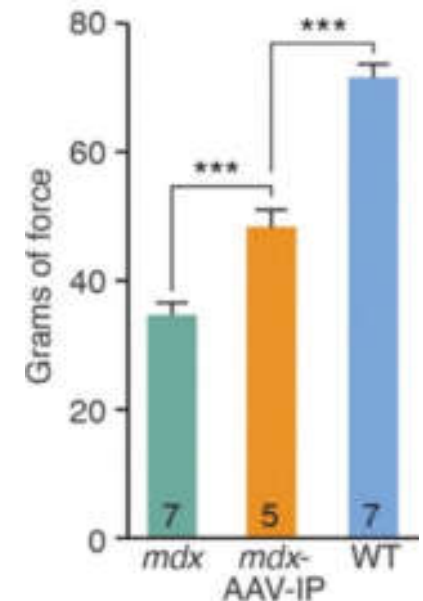
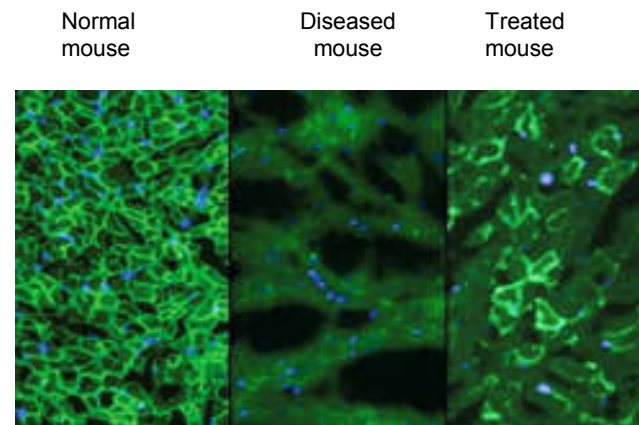
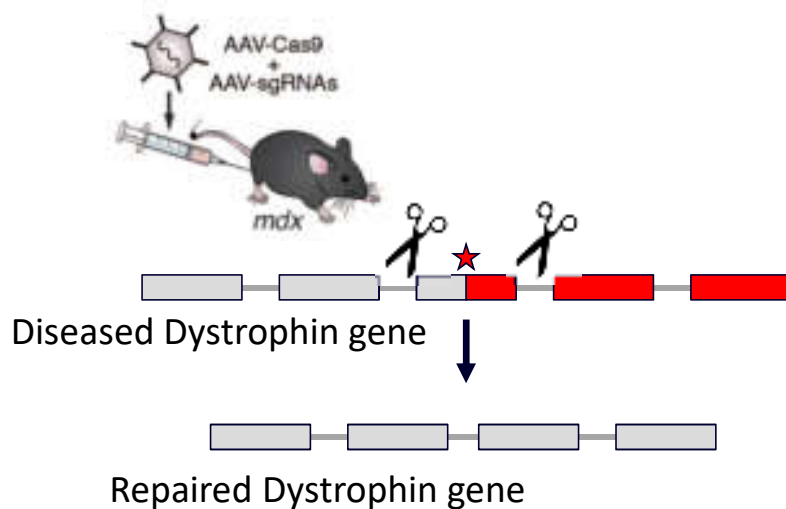
How cutting DNA can be used therapeutically



CRISPR is providing a therapeutic approach for muscular dystrophy in preclinical models

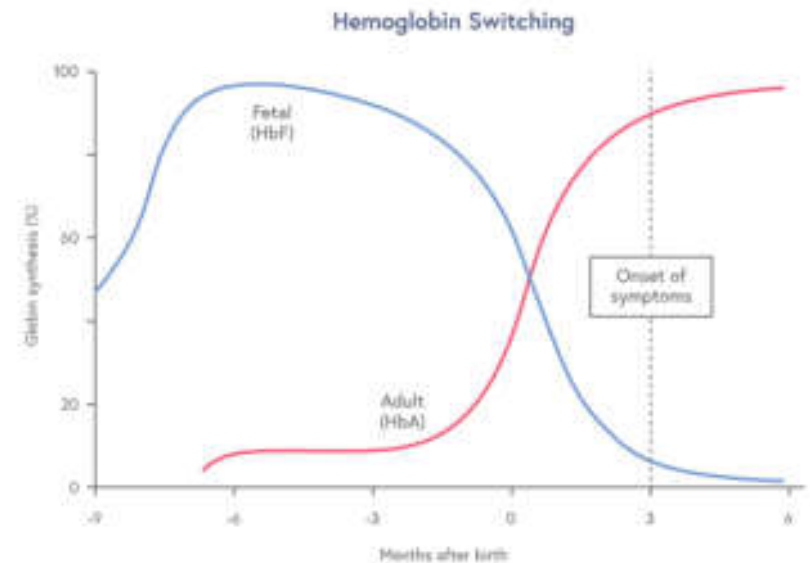
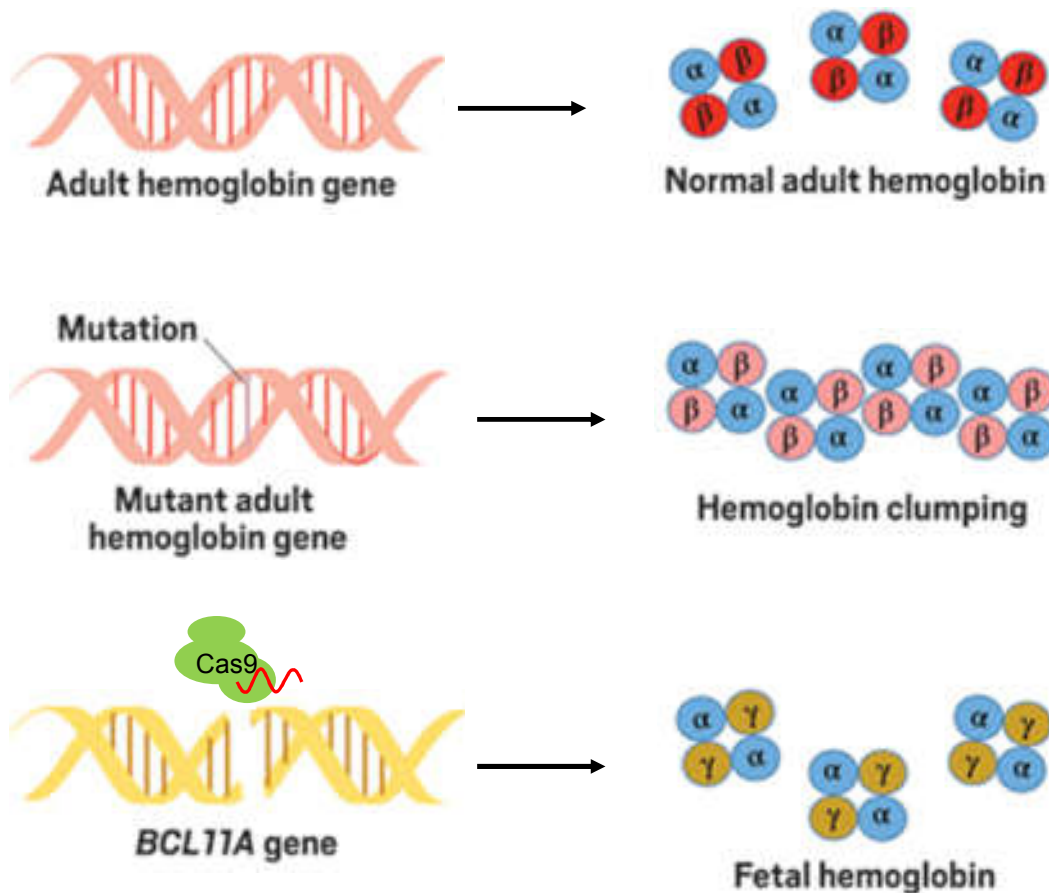


- Gene editing techniques using CRISPR-Cas9 have recently been used to correct this mutation in a mouse model



Long et al., Science 2016
Nelson et al., Science 2016

CRISPR is providing a therapeutic approach for haemoglobinopathies in human patients – Phase I/II



CRISPR gene editing in humans appears safe, and potentially effective

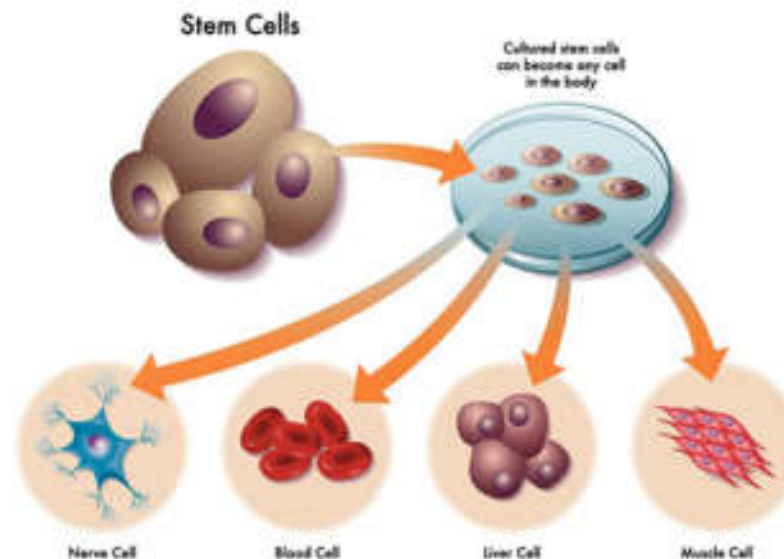
Crispr Therapeutics and Vertex Pharmaceuticals release first data from sickle cell disease and β -thalassaemia clinical trial

by Ryan Cross

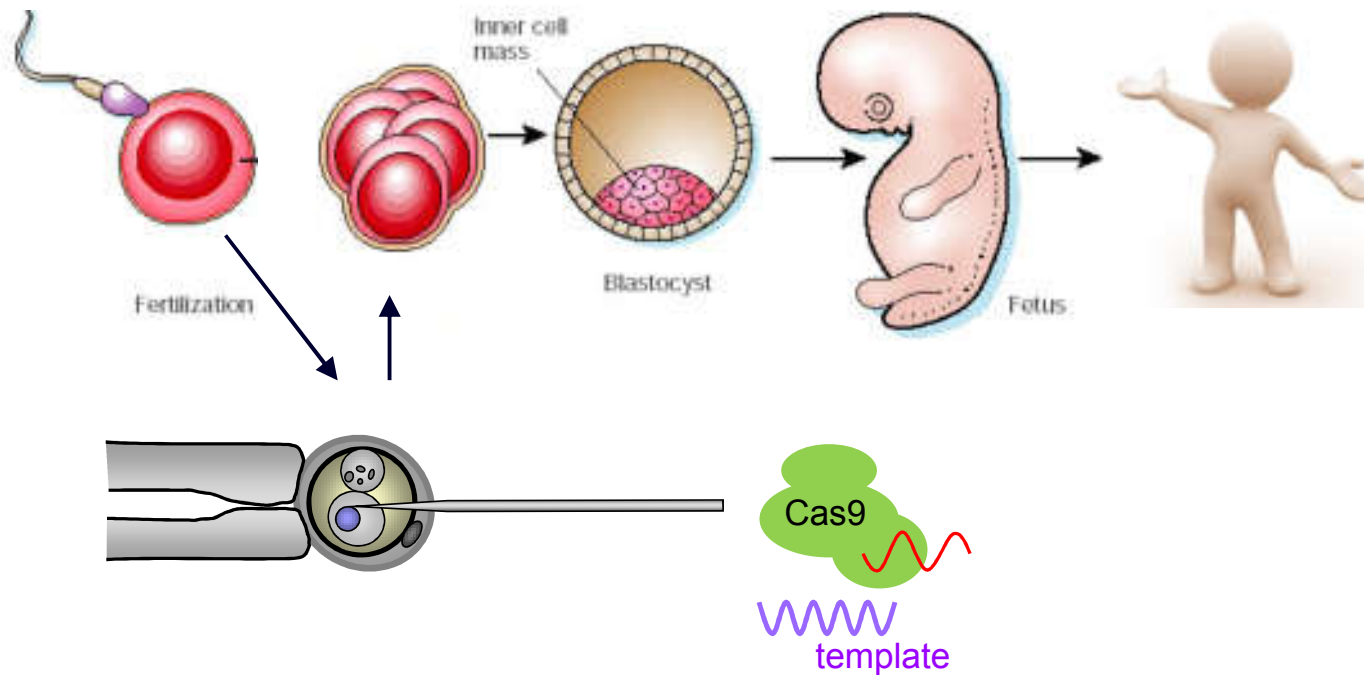
20TH MARCH 2021 (APPROVED BY VOLUME 37, ISSUE 48)

Which cells to treat?

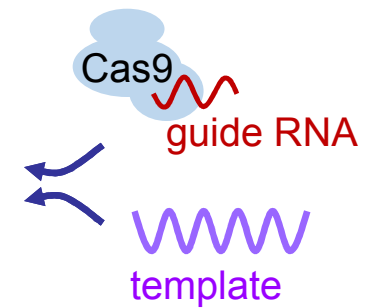
- A frequent problem with gene therapy is that the therapeutic gene delivery has to keep being applied to the diseased organ
- If, however, we target stem cells which can repopulate a particular tissue or organ, this could be a permanent fix.



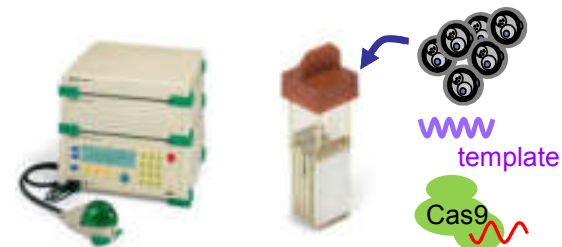
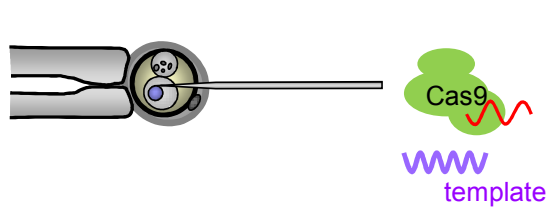
Can we / should we fix genes in the embryo?



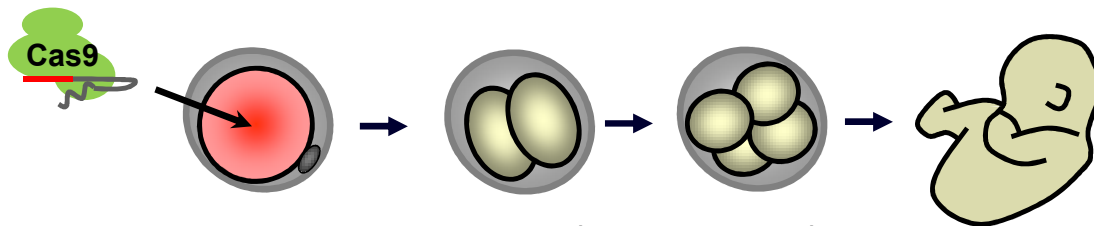
CRISPR/Cas9 Mutagenesis within the zygote



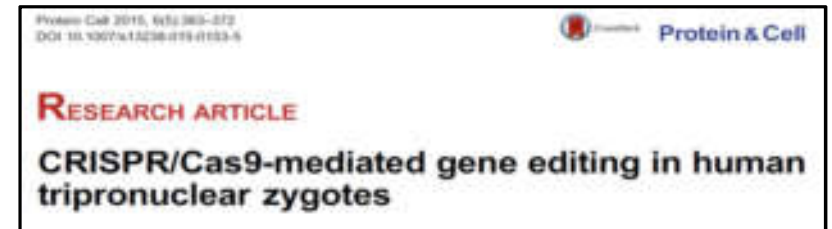
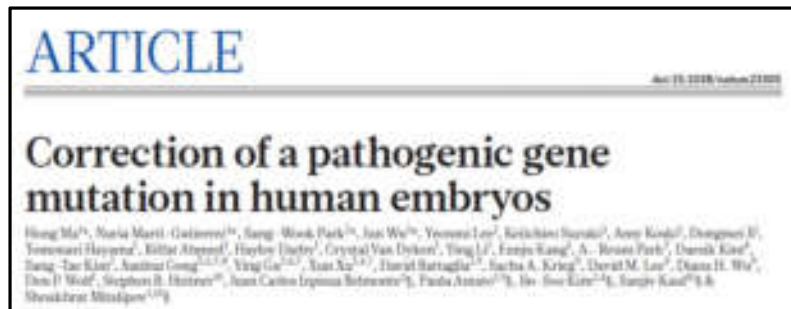
Alternative methods for delivery to the zygote



What works in mouse, works in human



- Disease correction in human embryos



Problems – off-target mutation

- The specificity of CRISPR is defined by only 20 nt
- (near) identical sequences may be present elsewhere
-and Cas9 nuclease tolerates certain mismatches
- How real is this problem when applying CRISPR/Cas9 nucleases in the single cell embryo



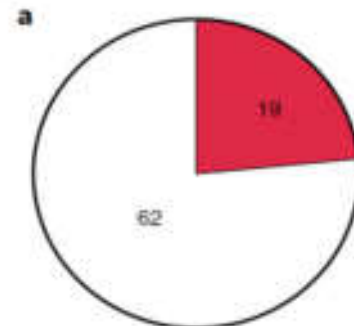
No unexpected CRISPR-Cas9 off-target activity revealed by trio sequencing of gene-edited mice

Vivek Iyer^{1*}, Katharina Boroviak^{2*}, Mark Thomas³, Brendan Doe⁴, Laura Riva⁵, Edward Ryder⁶, David J. Adams⁶

CRISPR off-target analysis in genetically engineered rats and mice

Keith R. Anderson¹, Maximilian Haessler², Colin Watanabe³, Vasantharajan Janakiraman⁴, Jessica Lund⁵, Zora Modrusan⁶, Jeremy Stinson⁷, Qixin Bei⁸, Andrew Buechler⁹, Charles Yu¹⁰, Sobha R. Thamminana¹¹, Lucinda Tam¹², Michael-Anne Sowick¹³, Tulja Alcantar¹⁴, Natasha O'Neill¹⁵, Jirjie Li¹⁶, Linda Tai¹⁷, Lisa Lima¹⁸, Merone Roosa-Girma¹⁹, Xin Rairdan²⁰, Steffen Durinck²¹ and Seren Warming^{22*}

■ Projects with OT
□ Projects without OT



Off-targets – solutions?



- Better algorithms predicting accuracy
- Engineered Cas9s with high accuracy
- Other CRISPR/Cas family members show higher levels of accuracy
- Accuracy is also vital for discriminating a mutant copy of a gene from a healthy copy

ARTICLE

doi:10.1038/s41467-018-01228-8

High-fidelity CRISPR–Cas9 nucleases with no detectable genome-wide off-target effects

Benjamin P. Kleinstiver^{1,2*}, Vikram Pattanajak^{1,2*}, Michelle S. Pres¹, Shengtao Q. Tian^{2,3}, Nho T. Nguyen¹, Zongli Zhang⁴ & J. Keith Joung^{1,2,5}

SpCas9-HF

Rationally engineered Cas9 nucleases with improved specificity

Ian M. Slaymaker^{1,2,3,4*}, Linyi Gao^{1,2*}, Bernd Zetsche^{1,2,3,4}, David A. Scott^{1,2,3,4}, Winston X. Yan^{1,2,3}, Feng Zhang^{1,2,3,4,5}

eSpCas9

Engineered CRISPR–Cas12a variants with increased activities and improved targeting ranges for gene, epigenetic and base editing

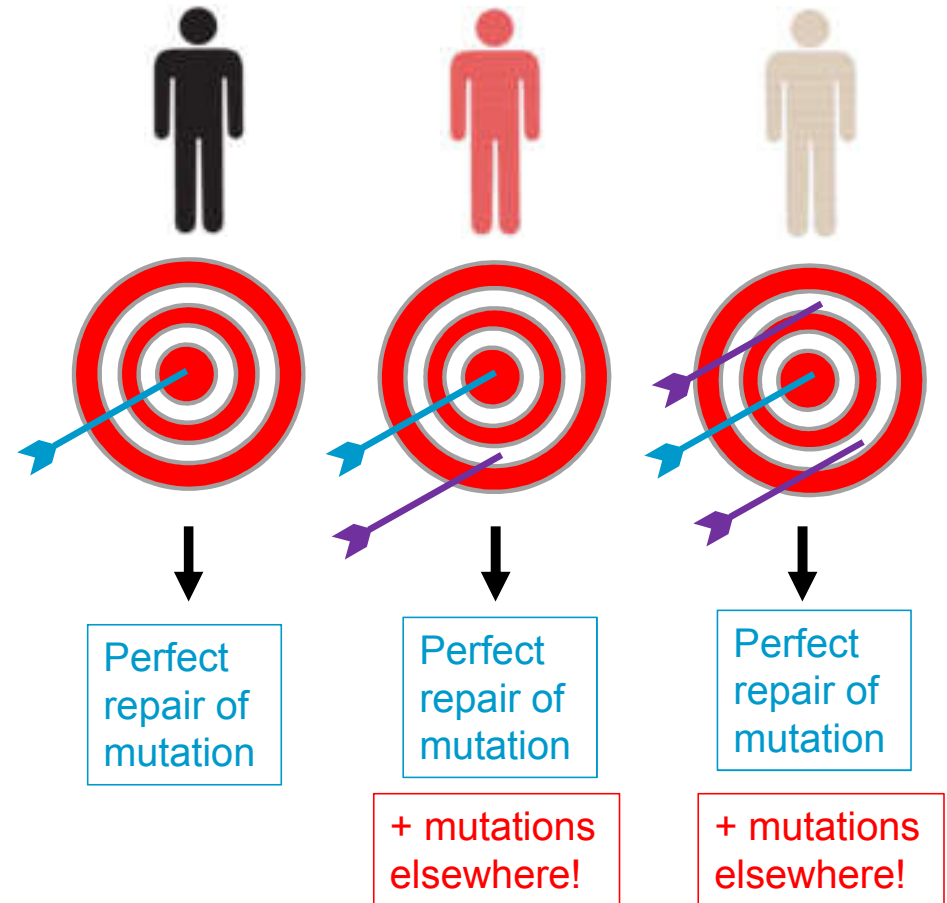
Benjamin P. Kleinstiver^{1,2,3,4*}, Alexander A. Sousa^{1,2,3,4}, Russell T. Walton^{1,2,3,4,5}, Y. Esther Tak^{1,2,3,4}, Jonathan Y. Hsu^{1,2,3}, Kendell Clement^{1,2,3,4}, Moira M. Welch^{1,2,3}, Joy E. Horng^{1,2,3}, Jose Malagon-Lopez^{1,2,3,4,5,6}, Irene Scarfo^{1,2,3}, Marcela V. Maus^{1,2,3}, Luca Pinello^{1,2,3,4}, Martin J. Aryee^{1,2,3,4,5} and J. Keith Joung^{1,2,3,4,5}

Real off-target prediction demands a personalized genome

Mutations found	Average per individual
Total variants	3,776,362
SNVs	3,579,423
Indels	196,940
Coding synonymous	11,742
Coding non-synonymous	11,468
Coding STOP/Splicing	478

Taliun et al bioRxiv2019

- Each of us differs on average at 1 nucleotide every 1000
- How can we predict off-target mutagenesis when we don't have the genome sequence of our patients



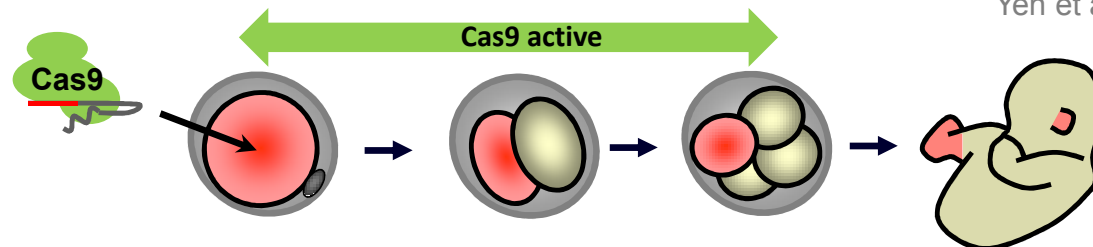
Problems – mosaicism



- Founder mice generated with CRISPR are almost invariably mosaic
- Not all cells are mutated/corrected!



Yen et al., 2014



Problems – large deletions and rearrangements



**nature
biotechnology**

Repair of double-strand breaks induced by CRISPR–Cas9 leads to large deletions and complex rearrangements

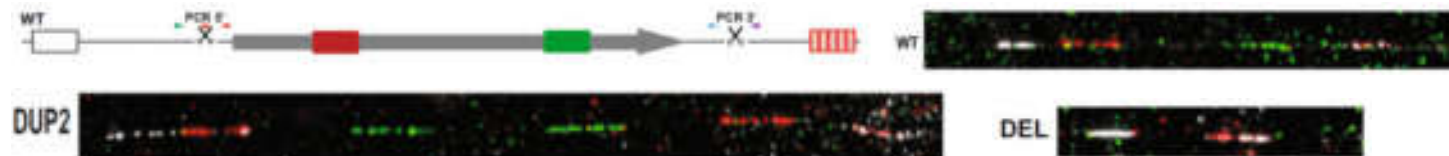
Michael Knicki, Kari Tienberg & Allan Bradley



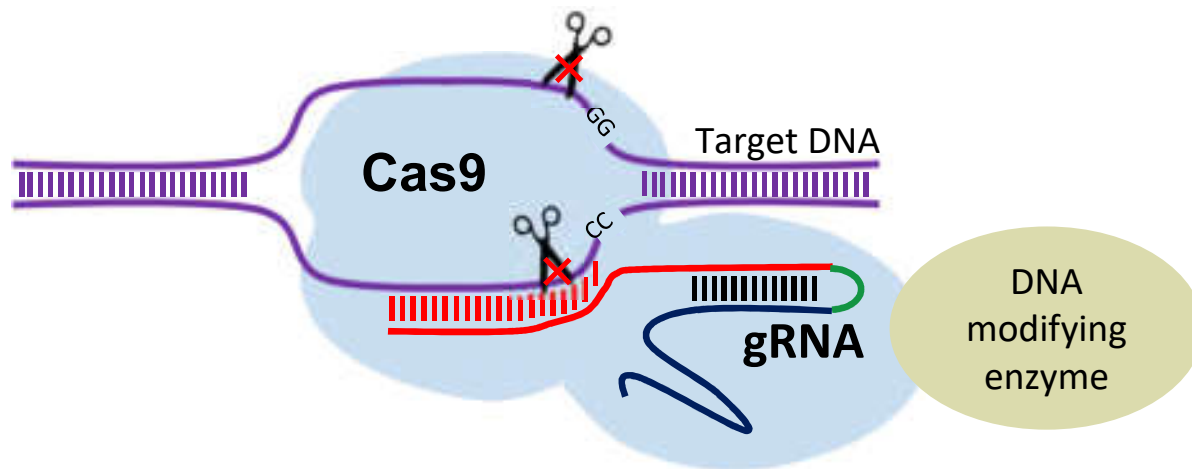
SCIENTIFIC REPORTS

OPEN Revealing hidden complexities of genomic rearrangements generated with Cas9

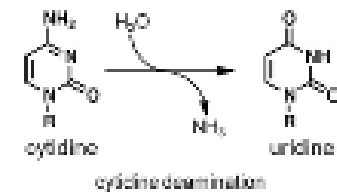
Received: 12 May 2017
Accepted: 10 September 2017
Katharina Benschke, Stefano Yu, Pengfeng Tang, Brandon Cole & Allan Bradley



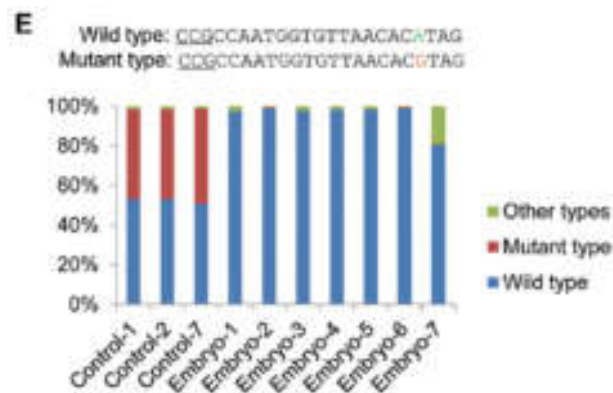
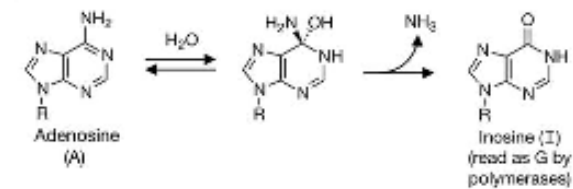
Solutions – avoid cutting the genome in the first place



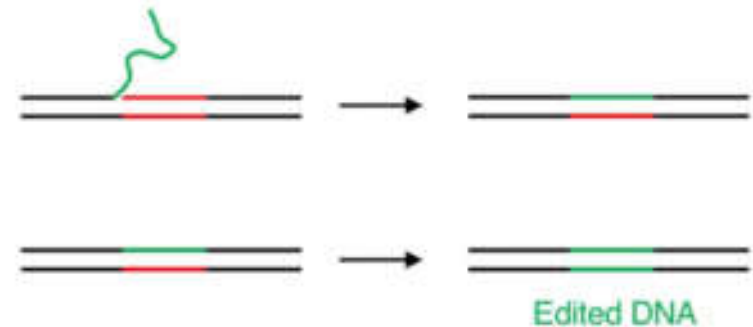
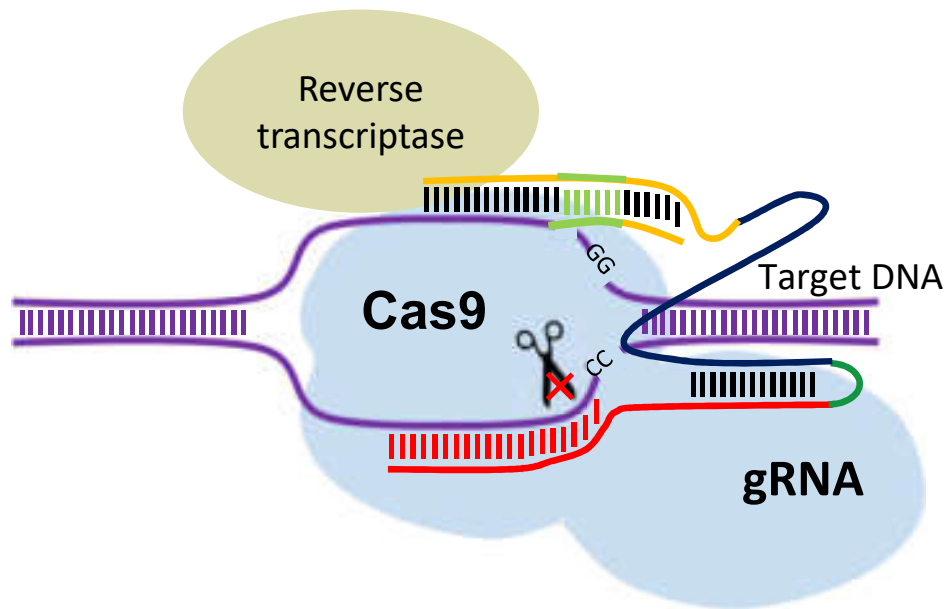
- C to T



- A to G



Prime editing – a new copy/paste technology



Article Search-and-replace genome editing without double-strand breaks or donor DNA

<https://doi.org/10.1038/s41586-019-1711-8>

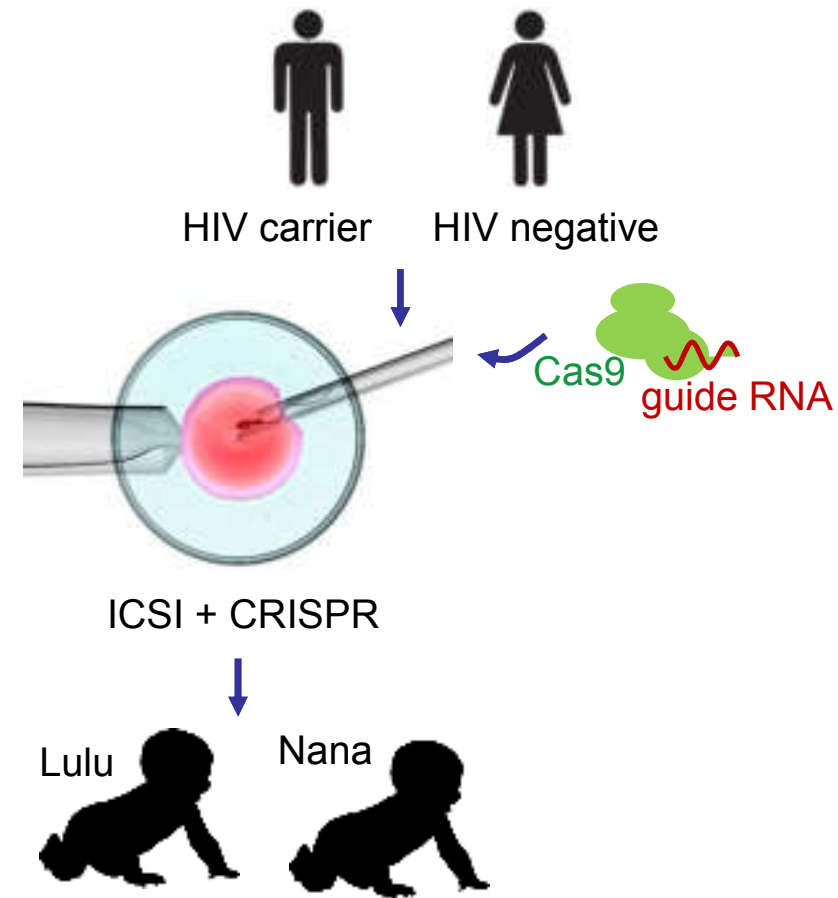
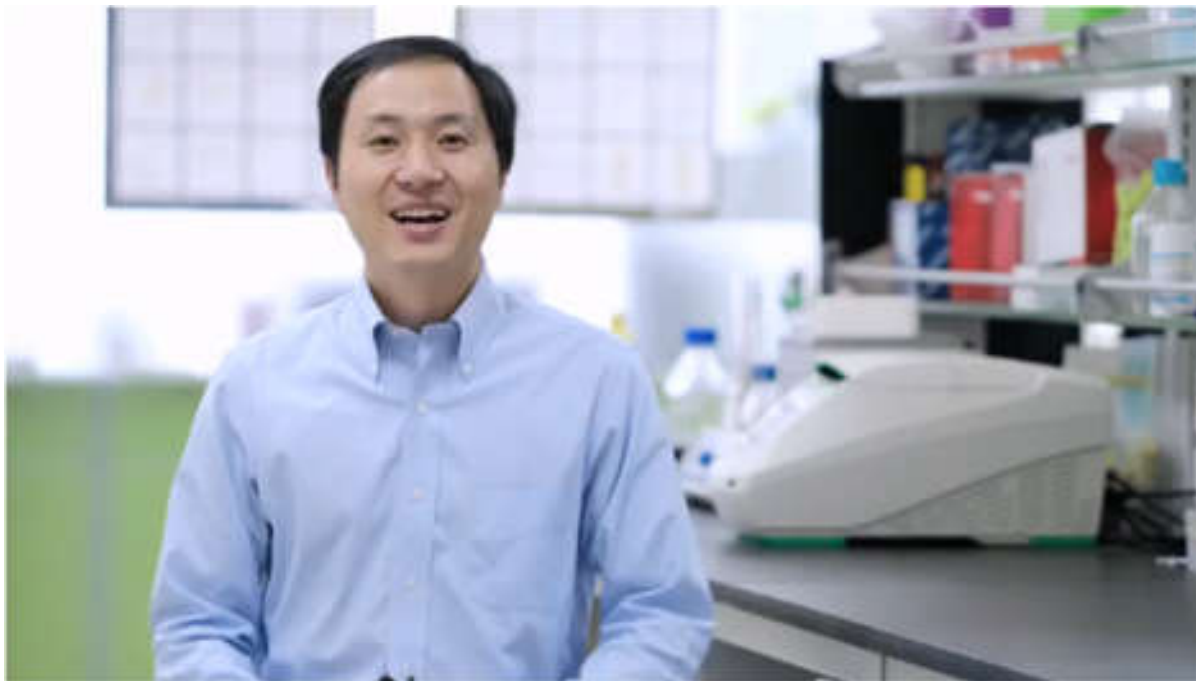
Received: 26 August 2019

Accepted: 10 October 2019

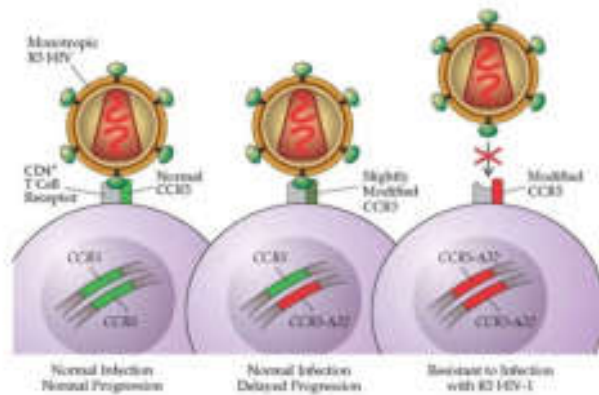
Andrew V. Anzalone^{1,2}, Peyton S. Randolph^{1,2}, Jessie R. Davis^{1,2}, Alexander A. Sousa^{1,2},
Luke W. Koblen^{1,2}, Jonathan M. Levy^{1,2}, Peter J. Chen^{1,2}, Christopher Wilson^{1,2},
Gregory A. Newby^{1,2,3}, Aditya Raguvaran^{1,2} & David R. Liu^{1,2,4}



Meet Dr. He Jiankui



Some people in the population are resistant to HIV



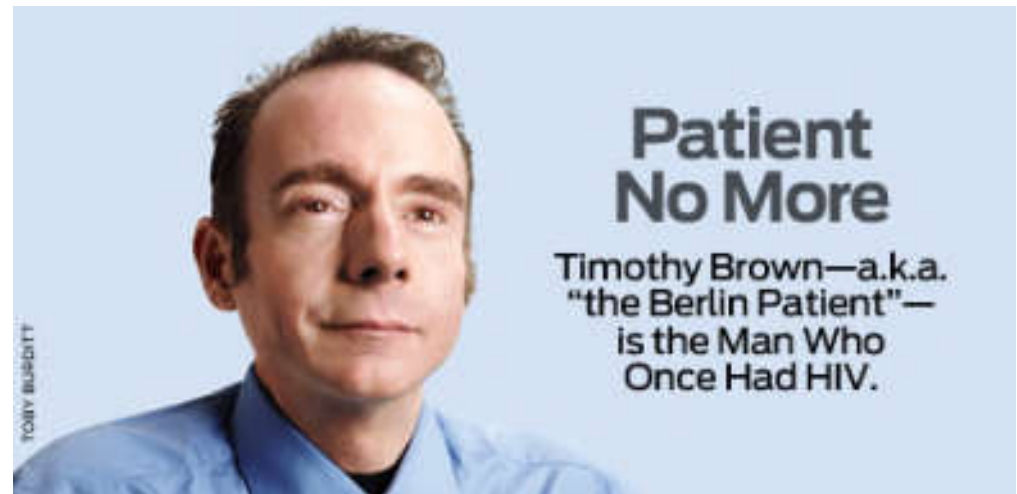
THE NEW ENGLAND JOURNAL OF MEDICINE

BRIEF REPORT

Long-Term Control of HIV by CCR5 Delta32/ Delta32 Stem-Cell Transplantation

Geno Hütter, M.D., Daniel Nowak, M.D., Maximilian Moosner, B.S.,
Susanne Gerngross, M.D., Anne Müllig, M.D., Kristina Allers, Ph.D.,
Thomas Schneider, M.D., Ph.D., Jörg Hofmann, Ph.D., Claudia Kücherer, M.D.,
Olga Blau, M.D., Igor W. Blau, M.D., Wolf K. Hofmann, M.D.,
and Eckhard Thiel, M.D.

- Naturally occurring mutation in a protein receptor, CCR5, stops HIV entry into cells
- If donor cells from a naturally HIV resistant patient are infused into an HIV patient, can they be cured?



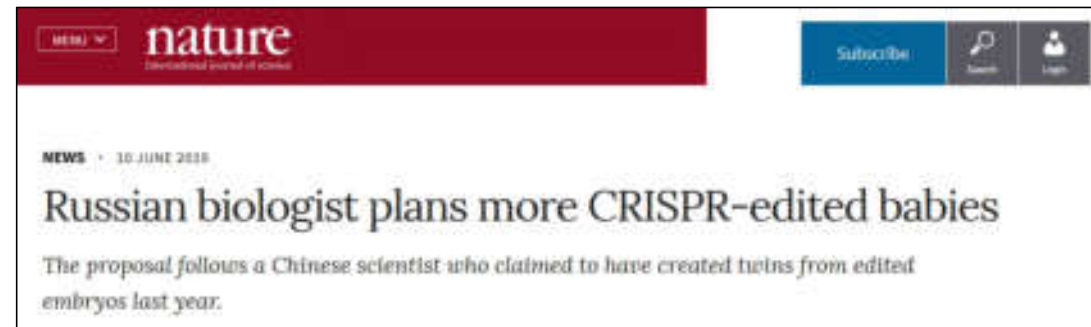
Dangerous and nonsensical use of the technology

- No clinical need for CCR5 gene editing
- The consequences of CCR5 knock-out throughout life in humans is unclear.
- CCR5 loss in mouse leads to increased susceptibility to other viral infections, such as influenza and West Nile virus.
- CCR5 loss has been associated with reduced longevity and altered response to neuronal injury



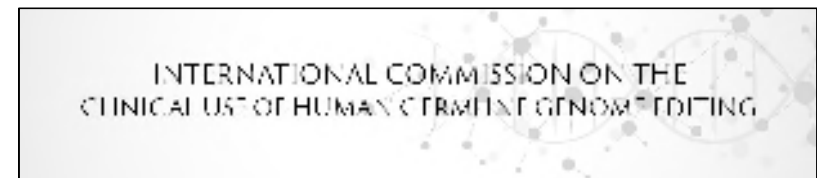
- Heterozygous embryo knowingly implanted – one normal copy of CCR5 remains.
- Biological activity of Nana's de novo alleles (15 bp deletion = 5 amino acids deletion) of CCR5 completely unclear.

A call for heightened regulation and a moratorium



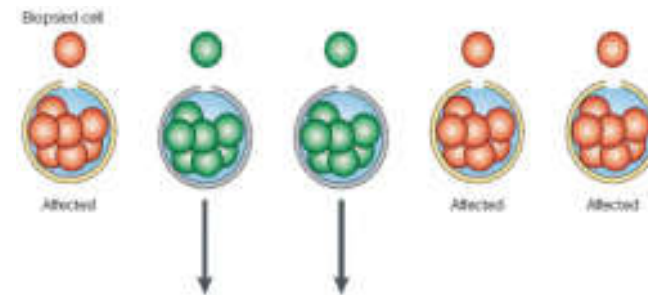
Statement of Principles on Genome Editing – August 27, 2019

“We assert that germline gene editing is currently inappropriate in human clinical settings”



But, is there a need for germline editing?

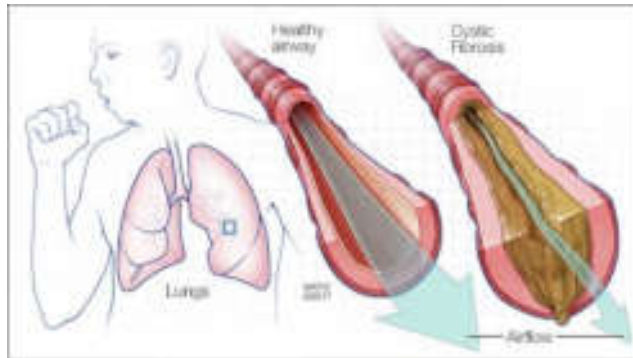
- Preimplantation Genetic Diagnosis (PGD) can be used to determine which embryos are healthy
- But PGD reduces the chances of success of ART
- The number of embryos obtained may be too low to allow the effective use of PGD
- Certain genetic situations aren't appropriate for PGD



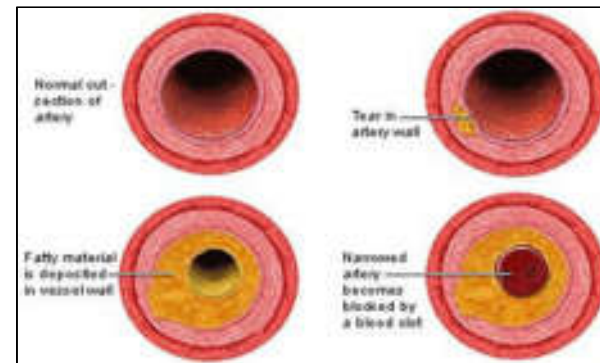
- Intentionally refraining from engaging in life-saving research is not morally defensible.
- Research is needed to ascertain and address any risks



What should we use genome editing for?



Genetic disease – which ones?



Reducing risk of heart / vascular disease



Perfect eyesight



Improved intelligence



Fitness and strength



Longevity

Summary



- Current status
 - CRISPR/Cas site-specific nuclease can be easily programmed to address virtually any genomic sequence, enabling gene editing
 - Their introduction to fertilized zygotes leads to efficient mutagenesis
 - Lack of precision and associated non-specific DNA damage is a concern
 - Next generation tools being rapidly developed to counter these concerns
- The near future
 - Greater understanding of DNA repair outcomes
 - Mining the bacterial kingdom to identify new enzymes and evolution of existing ones
 - More information about clinical trial safety where CRISPR is being used in somatic tissues ex vivo (and also now in vivo).
 - Optimization and thorough safety assessment of base editing
 - A therapeutic application in human embryos

Acknowledgements



Chris Preece

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Samy Alghadban

Amine Bouchareb



National Centre
for the Replacement
Refinement & Reduction
of Animals in Research